



AESA BASED IPM PACKAGE

AESA based IPM – Drumstick



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FOREWORD

Intensive agricultural practices relying heavily on chemical pesticides are a major cause of wide spread ecological imbalances resulting in serious problems of insecticide resistance, pest resurgence and pesticide residues. There is a growing awareness world over on the need for promoting environmentally sustainable agriculture practices.

Integrated Pest Management (IPM) is a globally accepted strategy for promoting sustainable agriculture. During last century, IPM relied substantially on economic threshold level and chemical pesticides driven approaches. However, since the late 1990s there is conscious shift to more ecologically sustainable Agro-Eco System Analysis (AESA) based IPM strategies. The AESA based IPM focuses on the relationship among various components of an agro-ecosystem with special focus on pest-defender dynamics, innate abilities of plant to compensate for the damages caused by the pests and the influence of abiotic factors on pest buildup. In addition, Ecological Engineering for pest management - a new paradigm to enhance the natural enemies of pests in an agro-ecosystem is being considered as an important strategy. The ecological approach stresses the need for relying on bio intensive strategies prior to use of chemical pesticides.

Sincere efforts have been made by resource personnel to incorporate ecologically based principles and field proven technologies for guidance of the extension officers to educate, motivate and guide the farmers to adopt AESA based IPM strategies, which are environmentally sustainable. I hope that the AESA based IPM packages will be relied upon by various stakeholders relating to Central and State government functionaries involved in extension and Scientists of SAUs and ICAR institutions in their endeavour to promote environmentally sustainable agriculture practices.

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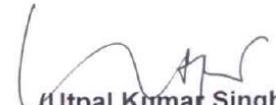
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FOREWORD

IPM is a holistic approach of crop protection based on the integration of multiple strategies viz., cultural, physical, mechanical, biological, botanicals and chemical. Over the years IPM underwent several changes, shifting its focus from damage boundary, economic injury to economic threshold. Currently most stake holders rely upon economic threshold levels (ETL) and tend to apply chemical pesticides at the first instance in the event of a pest attack, though Government of India has advocated need based and judicious application of chemicals. This approach is likely to cause adverse effects on agro-ecosystems and increase the cost of agricultural production due to problems of pest resurgence, insecticide resistance and sustainability.

During the late 90s FAO started advocating Agro-Ecosystem Analysis (AESA) based IPM. Experience in different countries have since shown that AESA, which takes into account ecological principles and relies on the balance that is maintained by biotic factors in an ecosystem has also resulted in reduction in cost of production and increase in yields. AESA based IPM also takes into account the need for active participation of farmers and promotes experiential learning and discovery based decision making by farmers. AESA based IPM in conjunction with ecological engineering for pest management promotes bio-intensive strategies as against current chemical intensive approaches, while retaining the option to apply chemical pesticides judiciously as a measure of last resort.

The resource persons of NIPHM and DPPQ&S have made sincere efforts in revising IPM packages for different crops by incorporating agro-ecosystem analysis, ecological engineering, pesticide application techniques and other IPM options with the active cooperation of crop based plant protection scientists working in State Agricultural Universities and ICAR institutions. I hope this IPM package will serve as a ready reference for extension functionaries of Central/ State Governments, NGOs and progressive farmers in adopting sustainable plant protection strategies by minimizing the dependence on chemical pesticides.


(Utpal Kumar Singh)



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PREFACE

Need for environmentally sustainable agricultural practices is recognised worldwide in view of the wide spread ecological imbalances caused by highly intensive agricultural systems. In order to address the adverse impacts of chemical pesticides on agro-ecosystems, Integrated Pest Management has evolved further from ETL based approach to Agro-ecosystem Analysis based Integrated Pest Management (IPM).

In AESA based IPM the whole agro-ecosystem, plant health at different stages, built-in-compensation abilities of the plant, pest and defender population dynamics, soil conditions, climatic factors and farmers' past experience are considered. In AESA, informed decisions are taken by farmers after field observation, AESA chart preparation followed by group discussion and decision making. Insect zoo is created to enable the farmer understand predation of pests by Natural Enemies. AESA based PHM also results in reduction of chemical pesticide usage and conserves the agro-ecosystems.

Ecological Engineering for Pest Management, a new paradigm, is gaining acceptance as a strategy for promoting Biointensive Integrated Pest Management. Ecological Engineering for Pest Management relies on cultural practices to effect habitat manipulation and enhance biological control. The strategies focus on pest management both below ground and above ground. There is a growing need to integrate AESA based IPM and principles of ecological engineering for pest management.

There is a rising public concern about the potential adverse effects of chemical pesticides on the human health, environment and biodiversity. The intensity of these negative externalities, though cannot be eliminated altogether, can be minimized through development, dissemination and promotion of sustainable biointensive approaches.

Directorate of Plant Protection Quarantine and Storage (DPPQS), has developed IPM package of practices during 2001 and 2002. These packages are currently providing guidance to the Extension Officers in transferring IPM strategies to farmers. These IPM package of practices, have been revised incorporating the principles of AESA based IPM in detail and also the concept of Ecological Engineering for Pest Management. It is hoped that the suggested practices, which aim at enhancing biodiversity, biointensive strategies for pest management and promotion of plant health, will enable the farmers to take informed decisions based on experiential learning and it will also result in use of chemical pesticides only as a last resort & in a safe and judicious manner.

(K. SATYAGOPAL)

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AESA BASED IPM PACKAGE FOR DRUMSTICK

Drumstick plant description:

Drumstick (*Moringa oleifera* Lam.; Family: Moringaceae) English common names include moringa, drumstick tree (from the appearance of the long, slender, triangular seed-pods), horseradish tree (from the taste of the roots, which resembles horseradish), ben oil tree or benzoil tree (from the oil which is derived from the seeds). It is a fast-growing, drought-resistant tree, native to the southern foothills of the Himalayas in northwestern India, and widely cultivated in tropical and sub-tropical areas where its young seed pods and leaves are used as a vegetable. *Moringa* derives from the Tamil word *murungai*.

India is the largest producer of moringa, with an annual production of 1.1 to 1.3 million tonnes of tender fruits. Among the states, Andhra Pradesh leads in both area and production followed by Karnataka and Tamil Nadu. Tamil Nadu is the pioneering state in so much as it has varied genotypes from diversified geographical areas and introductions from Sri Lanka. *Moringa* can be grown as an annual or perennial plant. In the first year all pods are edible. Later years also bear non edible bitter pods. Therefore *Moringa* is often commercially cultivated annually. On less favorable locations the perennial cultivation has big advantages. Erosion is much smaller with perennial cultivation. Perennial cultivation of *Moringa* is also practiced in agroforestry.

Moringa oleifera is a fast-growing, evergreen, deciduous tree. It can reach a height of 10-12 m and the trunk can reach a diameter of 45 cm. The bark has a whitish-grey colour and is surrounded by thick cork. Young shoots have purplish or greenish-white hairy bark. The tree has an open crown of drooping, fragile branches and the leaves build up feathery foliage of tripinnate leaves. The flowers are fragrant and bisexual, surrounded by five unequal thinly veined yellowish-white petals. The flowers are approximately 1-1.5 cm long and 2 cm broad. They grow on slender hairy stalks in spreading or drooping later flower clusters which have a longitude of 10–25 cm. Flowering begins within the first six months after planting. In seasonally cool regions, flowering will only occur once a year between April and June. In more constant seasonal temperature and with constant rainfall, flowering can happen twice or even all year-round. The fruit is a hanging, three-sided brown capsule of 20–45 cm size which holds dark brown, globular seeds with a diameter of approximately 1 cm. The seeds have three whitish papery wings and are dispersed by wind and water. In cultivation, it is often cut back annually to 1–2 meters and allowed to regrow so the pods and leaves remain within arm's reach.



I. PESTS

A. Pests of National Significance

1. Insect pests

Except bark eating caterpillar all the insect pests of drumstick reported here are specific to drumstick (monophagous) (Satti et al. 2013)

1.1 Moringa hairy caterpillar: *Eupterote mollifera* Walker (Lepidoptera: Eupterotidae)

1.2 Moringa budworm: *Noorda moringae* Walker (Lepidoptera: Pyraustidae)

1.3 Leaf caterpillar: *Noorda blitealis* Walker (Lepidoptera: Pyraustidae)

1.4 Pod fly: *Gitona distigma* Meigen (Diptera: Drosophilidae)

1.5 Bark eating caterpillar: *Indarbela tetraonis* Moore (Lepidoptera: Cossidae)

2. Diseases

2.1 Damping off: *Pythium aphanidermatum* (Edson) Fitzp, *P. debaryanum* R. Hesse, and *Rhizoctonia solani* J.G. Kühn

2.2 Twig canker: *Fusarium pallidoroseum* (Cooke) Sacc

3. Weeds

3.1 Major *Kharif* weeds

Broadleaf

3.1.1 Pigweed: *Amaranthus viridis* Hook. F. (Amaranthaceae)

3.1.2 Swine cress: *Coronopus didymus* (L.) Sm. (Brassicaceae)

3.1.3 Black nightshade: *Solanum nigrum* L. (Solanaceae)

3.1.4 Common purselane: *Portulaca oleracea* L. (Portulacaceae)

3.1.5 False amaranth: *Digera arvensis* Forssk. (Amaranthaceae)

Grasses

3.1.6 Rabbit/crow foot grass: *Dactyloctenium aegyptium* (L.) Beauv. (Poaceae)

3.1.7 Crab grass: *Digiteria sanguinalis* (L.) Willd. (Poaceae)

3.1.8 Barnyard grass: *Echinochloa crusgalli* (L.) Scop. (Poaceae)

Sedges

3.1.9 Purple nutsedge: *Cyperus rotundus* L. (Cyperaceae)

3.1.10 Flat sedge: *Cyperus iria* L. (Cyperaceae)

5.1 Major Rabi weeds

Broadleaf

3.2.1 Lamb's quarter: *Chenopodium album* L. (Chenopodiaceae)

3.2.2 Scarlet pimpernel: *Anagallis arvensis* L. (Primulaceae)

3.2.3 Sweet clover: *Melilotus indica* (L.) All. (Fabaceae)

3.2.4 Fine leaf fumitory: *Fumaria parviflora* Lam. (Fumariaceae)

3.2.5. Corn spurry: *Spergula arvensis* L. (Caryophylliaceae)

Grasses

3.2.6 Blue grass: *Poa annua* L. (Poaceae)

3.2.7 Canary grass: *Phalaris minor* Retz. (Poaceae)

II. AGRO-ECOSYSTEM ANALYSIS (AESAs) BASED INTEGRATED PEST MANAGEMENT (IPM)

A. AESA

The IPM has been evolving over the decades to address the deleterious impacts of synthetic chemical pesticides on environment ultimately affecting the interests of the farmers. The economic threshold level (ETL) was the basis for several decades but in modern IPM (FAO 2002) emphasis is given to AESA where farmers take decisions based on larger range of field observations. The health of a plant is determined by its environment which includes physical factors (i.e. soil, rain, wind, sunshine hours etc.) and biological factors (i.e. pests, diseases and weeds). All these factors can play a role in the balance which exists between herbivore insects and their natural enemies. Understanding the intricate interactions in an ecosystem can play a critical role in pest management.

Decision making in pest management requires a thorough analysis of the agro-ecosystem. Farmer has to learn how to observe the crop, how to analyze the field situation and how to make proper decisions for their crop management. This process is called the AESA. Participants of AESA will have to make a drawing on a large piece of paper (60 x 80 cm), to include all their observations. The advantage of using a drawing is that it forces the participants/farmers to observe closely and intensively. It is a focal point for the analysis and for the discussions that follow, and the drawing can be kept as a record.

AESA is an approach, which can be gainfully employed by extension functionaries and farmers to analyze the field situations with regards to pests, defenders, soil conditions, plant health and the influence of climatic factors and their relationship for growing a healthy crop. The basic components of AESA are:

- Plant health at different stages
- Built-in compensation abilities of plants
- Pest and defender population dynamics
- Soil conditions
- Climatic factors
- Farmers past experience

Principles of AESA based IPM:

Grow a healthy crop

- Select a variety resistant/tolerant to major pests
- Select healthy seeds/seedlings/planting material
- Treat the seeds/seedlings/planting material with recommended pesticides especially biopesticides
- Follow proper spacing
- Soil health improvement (mulching and green manuring wherever applicable)
- Nutrient management especially organic manures and biofertilizers based on the soil test results. If the dosage of nitrogenous fertilizers is too high the crop becomes too succulent and therefore susceptible to insects and diseases. If the dosage is too low, the crop growth is retarded. So, the farmers should apply an adequate amount for best results. The phosphatic fertilizers should not be applied each and every season as the residual phosphate of the previous season will be available for the current season also.
- Proper irrigation

Observe the field regularly (climatic factors, soil and biotic factors)

Farmers should

- Monitor the field situation at least once a week (soil, water, plants, pests, natural enemies, weather factors etc.)
- Make decisions based on the field situation and P: D ratio
- Take direct action when needed (e.g. collect egg masses, remove infested plants etc.)



Plant compensation ability

Compensation is defined as the replacement of plant biomass lost to herbivores and has been associated with increased photosynthetic rates and mobilization of stored resources from source organs to sinks (e.g., from roots and remaining leaves to new leaves) during active vegetative growth period. Plant tolerance to herbivory can arise from the interaction of a variety of plant traits and external environmental factors. Several studies have documented such compensation through increased growth and photosynthetic rate.

Understand and conserve defenders

- Know defenders/natural enemies to understand their role through regular observations of the agro-ecosystem
- Avoid the use of chemical pesticides especially with broad-spectrum activity

Insect zoo

In field various types of insects are present. Some are beneficial and some may be harmful. Generally farmers are not aware about it. Predators (friends of the farmers) which feed on pests are not easy to observe in crop field. Insect zoo concept can be helpful to enhance farmers' skill to identify beneficial and harmful insects. In this method, unfamiliar/unknown predators are collected in plastic containers with brush from the field and brought to a place for study. Each predator is placed inside a plastic bottle together with parts of the plant and some known insect pests. Insects in the bottle are observed for certain time and determined whether the test insect is a pest (feeds on plant) or a predator (feeds on other insects).

Pest: Defender ratio (P: D ratio):

Identifying the number of pests and beneficial insects helps the farmers to make appropriate pest management decisions. Sweep net, visual counts etc. can be adopted to arrive at the numbers of pests and defenders. The P: D ratio can vary depending on the feeding potential of natural enemy as well as the type of pest. The natural enemies of drumstick pests can be divided into 1. parasitoids; 2. predators; and 3. pathogens.

Model agro-ecosystem analysis chart

Date:
Village:
Farmer:



	Decision taken based on the analysis of field situations
Soil conditions	:
Weather conditions	:
Diseases types and severity	:
Weeds types and intensity	:
Rodent damage (if any)	:
No. of insect pests	:
No. of natural enemies	:
P: D ratio	:

The general rule to be adopted for management decisions relying on the P: D ratio is 2:1. However, some of the parasitoids and predators will be able to control more than 2 pests. Wherever specific P: D ratios are not found, it is safer to adopt the 2: 1, as P: D ratio. Whenever the P: D ratio is found to be favourable, there is no need for adoption of other management strategies. In cases where the P: D ratio is found to be unfavourable, the farmers can be advised to resort to inundative release of parasitoids/predators depending upon the type of pest. In addition to inundative release of parasitoids and predators, the usage of microbial biopesticides and biochemical biopesticides such as insect growth regulators, botanicals etc. can be relied upon before resorting to synthetic chemical pesticides.

Decision making

Farmers become experts in crop management

Farmers have to make timely decisions about the management of their crops. AESA farmers have learned to make these decisions based on observations and analysis viz. abiotic and biotic factors of the crop ecosystem. The past experience of the farmers should also be considered for decision making. However, as field conditions continue to change and new technologies become available, farmers need to continue improving their skills and knowledge.

- Farmers are capable of improving farming practices by experimentation
- Farmers can share their knowledge with other farmers

AESA methodology

- Go to the field in groups (about 5 farmers per group). Walk across the field and choose 20 plants/ acre randomly. Observe keenly each of these plants and record your observations:
 - Plant: Observe the plant height, number of branches, crop stage, deficiency symptoms etc.
 - Pests: Observe and count pests at different places on the plant.
 - Defenders (natural enemies): Observe and count parasitoids and predators.
 - Diseases: Observe leaves and stems and identify any visible disease symptoms and severity.
 - Rats: Count number of plants affected by rats.
 - Weeds: Observe weeds in the field and their intensity.
 - Water: Observe the water situation of the field.
 - Weather: Observe the weather condition.

- While walking in the field, manually collect insects in plastic bags. Use a sweep net to collect additional insects. Collect plant parts with disease symptoms.
- Find a shady place to sit as a group in a small circle for drawing and discussion.
- If needed, kill the insects with some chloroform (if available) on a piece of cotton.
- Each group will first identify the pests, defenders and diseases collected.
- Each group will then analyze the field situation in detail and present their observations and analysis in a drawing (the AESA drawing).
- Each drawing will show a plant representing the field situation. The weather condition, water level, disease symptoms, etc. will be shown in the drawing. Pest insects will be drawn on one side. Defenders (beneficial insects) will be drawn on another side. Write the number next to each insect. Indicate the plant part where the pests and defenders were found. Try to show the interaction between pests and defenders.
- Each group will discuss the situation and make a crop management recommendation.
- The small groups then join each other and a member of each group will now present their analysis in front of all participants.
- The facilitator will facilitate the discussion by asking guiding questions and makes sure that all participants (also shy or illiterate persons) are actively involved in this process.
- Formulate a common conclusion. The whole group should support the decision on what field management is required in the AESA plot.
- Make sure that the required activities (based on the decision) will be carried out.
- Keep the drawing for comparison purpose in the following weeks.

Data recording

Farmers should record data in a notebook and drawing on a chart

- Keeping records of what has happened help us making an analysis and draw conclusions

Data to be recorded

- **Plant growth (weekly):** Height of plant; number of leaves
- **Crop situation (e.g. for AESA):** Plant health; pests, diseases, weeds; natural enemies; soil condition; irrigation; weather conditions
- **Input costs:** Seeds; fertilizer; pesticides; labour
- **Harvest:** Yield (Kg/acre); price of produce (Rs./Kg)
- Price of produce (Rs./Kg)

Some questions that can be used during the discussion

- Summarize the present situation of the field?
- What crop management aspect is most important at this moment?
- Is there a big change in crop situation compared to last visit? What kind of change?
- Is there any serious pest or disease outbreak?
- What is the situation of the beneficial insects?
- Is there a balance in the field between pests and defenders?
- Were you able to identify all pests and diseases?
- Do you think the crop is healthy?
- What management practices are needed at this moment?

- When will it be done? Who will do it? Make sure that responsibilities for all activities are being discussed.
- Are you expecting any problems to emerge during the coming week such as congenial weather conditions for pest buildup?
- What problems? How can we avoid it? How can we be prepared?
- Summarize the actions to be taken.



Advantages of AESA over ETL

One of the problems of the ETL is that it is based on parameters that are changing all the time, and that are often not known. The damage or losses caused by a certain density of insects cannot be predicted at all. In ETL the due recognition of the role of natural enemies in decreasing pest population is ignored. Farmers cannot base their decisions on just a simple count of pests. They will have to consider many other aspects of the crop (crop ecology, growth stage, natural enemies, weather condition, etc.) and their own economic and social situation before they can make the right crop management decisions. In ETL based IPM, natural enemies, plant compensation ability and abiotic factors are not considered. In AESA based IPM emphasis is given to natural enemies, plant compensation ability, abiotic factors and P: D ratio.

AESA and farmer field school (FFS)

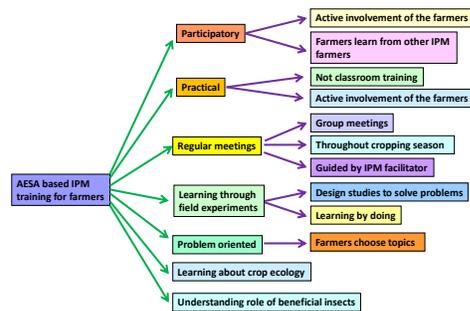
AESA is a season-long training activity that takes place in the farmer field. It is season-long so that it covers all the different developmental stages of the crop and their related management practices. The process is always learner-centered, participatory and relying on an experiential learning approach and therefore it has become an integral part of FFS.

Farmers can learn from AESA

- Identification of pests and their nature of damage
- Identification of natural enemies
- Management of pests
- Water and nutrient management
- Influence of weather factors on pest buildup
- Role of natural enemies in pest management



FFS to teach AESA based IPM skills



B. Field scouting

AESA requires skill. So only the trained farmers can undertake their exercise. However, other farmers also can do field scouting in their own fields at regular intervals to monitor the major pest situation.

Surveillance on pest occurrence at the main field should commence soon after crop establishment after transplanting and at weekly intervals thereafter. In each of the fields, select five spots randomly. Select five random plants at each spot for recording counts of insects as per procedure finalized for individual insects.

For insect pests:

Bud worm and pod fly larvae: Count the total number of fruiting parts checked. Count the total number of fruiting parts with bud worm or pod fly damage (calculate the percentage damaged fruiting parts).

For diseases:

Whenever scouting, be aware that symptoms of plant disease problems may be caused by any biotic factors such as fungal, bacterial, viral pathogens or abiotic factors such as weather, fertilizers, nutrient deficiencies, pesticides and abiotic soil problems. In many cases, the cause of the symptom is not obvious. Close examination, and laboratory culture and analysis are required for proper diagnosis of the causal agent of disease. Generally fungal diseases cause the obvious symptoms with irregular growth, pattern & colour (except viruses), however abiotic problems cause regular, uniform symptoms. Pathogen presence (signs) on the symptoms can also be observed like fungal growth, bacterial ooze etc. Specific and characteristic symptoms of the important plant diseases are given in description of diseases section.

Root sampling: Always check plants that appear unhealthy. If there are no obvious symptoms on plants, examine plants randomly and look for lesions or rots on roots and stems. Observe the signs of the causal organism (fungal growth or ooze). It is often necessary to wash the roots with water to examine them properly. If the roots are well developed, cut them to examine the roots for internal infections (discolouration & signs). Count the total number of roots damaged/infested/infected due to rot should be counted and incidence should be recorded.

Leaf sampling: Examine all leaves of each plant for lesions. Leaf diseases cause most damage during the seedling and flowering stages of plant growth. Observe for the symptoms and signs

on the infected plant parts. Determine the percent area of leaf infection by counting the number of leaves (leaf area diameter)/plant infected due to disease and incidence should be recorded.

Stem, flower and fruit sampling: Carefully examine the stem, flower, and fruit of plants for symptoms and signs of fungal or bacterial diseases. The stem, flower, and fruit should be split or taken apart and examined for discoloration caused by fungi and bacteria. Count the number of stems, flowers and fruits infected due to disease and percent disease incidence should be recorded.

C. Light traps

Set up light trap @ 1 trap/acre at mid canopy level for monitoring and mass trapping insects. Light traps with exit option for natural enemies of smaller size should be installed and operate around the dusk time (6 pm to 10 pm).

III. ECOLOGICAL ENGINEERING FOR PEST MANAGEMENT

Ecological engineering for pest management has recently emerged as a paradigm for considering pest management approaches that rely on the use of cultural techniques to effect habitat manipulation and to enhance biological control. The cultural practices are informed by ecological knowledge rather than on high technology approaches such as synthetic pesticides and genetically engineered crops (Gurr et al. 2004).

Natural enemies may require

1. Food in the form of pollen and nectar for adult natural enemies.
2. Shelter such as overwintering sites, moderate microclimate, etc.
3. Alternate host when primary host are not present.

Ecological engineering for pest management – Above ground:

- Raise the flowering plants / compatible cash crops along the field border by arranging shorter plants towards main crop and taller plants towards the border to attract natural enemies as well as to avoid immigrating pest population
- Grow flowering plants on the internal bunds inside the field
- Not to uproot weed plants those are growing naturally such as *Tridax procumbens*, *Ageratum* sp, *Alternanthera* sp etc. which act as nectar source for natural enemies
- Not to apply broad spectrum chemical pesticides, when the P: D ratio is favourable. The plant compensation ability should also be considered before applying chemical pesticides.

Ecological engineering for pest management – Below ground:

- Crop rotations with leguminous plants which enhance nitrogen content.
- Keep soils covered year-round with living vegetation and/or crop residue.
- Add organic matter in the form of farm yard manure (FYM), vermicompost, crop residue which enhance below ground biodiversity.
- Reduce tillage intensity so that hibernating natural enemies can be saved.
- Apply balanced dose of nutrients using biofertilizers.
- Apply mycorrhiza and plant growth promoting rhizobacteria (PGPR)
- Apply *Trichoderma* spp. and *Pseudomonas fluorescens* as seed/seedling/planting material, nursery treatment and soil application (if commercial products are used, check

for label claim. However, biopesticides produced by farmers for own consumption in their fields, registration is not required).

Due to enhancement of biodiversity by the flowering plants, parasitoids and predators (natural enemies) number also will increase due to availability of nectar, pollen, fruits, insects, etc. The major predators are a wide variety of spiders, ladybird beetles, long horned grasshoppers, *Chrysoperla*, earwigs, etc.

IV. CROP STAGE-WISE IPM

Management	Activity
Pre-sowing*	
	<p><u>Common cultural practices:</u></p> <ul style="list-style-type: none"> • Deep summer ploughing • Destroy the alternate host plants • Avoid planting overlapping crops in adjacent area.
Nutrients	<ul style="list-style-type: none"> • Apply FYM or compost @ 7-8 Kg per pit. • In addition, basal dose of N: P: K in 350: 100: 200 g/pit should also be incorporated at the time of planting.
Weeds	<ul style="list-style-type: none"> • Remove or incorporate previous crop residues before planting. • Plan to grow suitable intercrops like legumes, ginger, tapioca etc.
Soil borne fungus, resting stages of pests	<ul style="list-style-type: none"> • Soil solarization: Cover the beds with polythene sheet of 45 gauge (0.45 mm) thickness for three weeks before sowing for soil solarization which will help in reducing the soil borne pests.
Damping off	<p><u>Cultural control:</u></p> <ul style="list-style-type: none"> • Raise nursery in light soil with proper drainage • Burning farm trash on the surface of the beds. • Sowing seed on raised beds of 6-8" high (15 cm). Use low seed rate of 650 g/40 sq mt.
<p>* Apply <i>Trichoderma viride/harzianum</i> and <i>Pseudomonas fluorescens</i> as seed/seedling/planting material, nursery treatment and soil application (if commercial products are used, check for label claim. However, biopesticides produced by farmers for own consumption in their fields, registration is not required).</p>	
Vegetative stage	
	<p><u>Common cultural practices:</u></p> <ul style="list-style-type: none"> • Collect and destroy crop debris • Collect and destroy disease infected and insect damaged plant parts • Provide irrigation at critical stages of the crop • Avoid water logging • Avoid water stress during flowering stage • Judicious use of fertilizers • Enhance parasitic activity by avoiding chemical spray, when 1-2 larval parasitoids are observed • Field sanitation <p><u>Common mechanical practices:</u></p> <ul style="list-style-type: none"> • Collection and destruction of eggs and early stage larvae • Handpick the older larvae during early stages (do not handpick hairy caterpillars with urticating hairs) • The infested shoots and seed capsules may be collected and destroyed • Handpick the gregarious caterpillars (do not handpick hairy caterpillars with urticating hairs) and the cocoons which are found on stem and destroy them in kerosene mixed water. • Use light trap @ 1/acre and operate between 6 pm and 10 pm • Erecting of bird perches @ 20/acre for encouraging predatory

	<p>birds such as King crow, common mynah etc.</p> <ul style="list-style-type: none"> • Set up bonfire during evening hours at 7-8 pm <p><u>Common biological practices:</u></p> <ul style="list-style-type: none"> • Conserve natural enemies through ecological engineering • Augmentative release of natural enemies
Nutrients	<ul style="list-style-type: none"> • Under low fertile soils top dressing of N @ 100 g/pit by soil incorporation should be done. • Micronutrient deficiency should be corrected by foliar spray of particular nutrient.
Weeds	<ul style="list-style-type: none"> • Crop should be weed free up to 3 months by interculture and hand weeding.
Hairy caterpillar	<ul style="list-style-type: none"> • Follow common cultural, mechanical and biological practices <p><u>Mechanical control:</u></p> <ul style="list-style-type: none"> • The larvae in groups on tree trunks killed by a burning torch/flame thrower
Budworm Bark caterpillar	<ul style="list-style-type: none"> • Follow common cultural, mechanical and biological practices
Reproductive stage	
Nutrients	<ul style="list-style-type: none"> • Micronutrient deficiency should be corrected by foliar spray of particular nutrient.
Weeds	<ul style="list-style-type: none"> • Remove all the weeds before seed formation to avoid further spread of weed seeds.
Pod fly	<p><u>Cultural control:</u></p> <ul style="list-style-type: none"> • Collect and destroy all the fallen and damaged fruits • Trap the adults using attractants such as citronella oil, eucalyptus oil, vinegar (acetic acid), dextrose or lactic acid • Rake up the soil under the trees or plough the infested field to destroy puparia. <p><u>Biological control</u></p> <ul style="list-style-type: none"> • Spray NSKE 5% during 50% fruit set and 35 days later

Note: The pesticide dosages and spray fluid volumes are based on high volume spray.

V. INSECTICIDE RESISTANCE AND ITS MANAGEMENT

Insecticide resistance: Resistance to insecticides may be defined as 'a heritable change in the sensitivity of a pest population that is reflected in the repeated failure of a product to achieve the expected level of control when used according to the label recommendation for that pest species' (IRAC). Cross-resistance occurs when resistance to one insecticide confers resistance to another insecticide, even where the insect has not been exposed to the latter product.

Causes of resistance development: The causes and rate at which insecticide resistance develops depend on several factors, including how rapidly the insects reproduce, the insects' level of resistance, the migration and host range of the insects, the insecticide's persistence and

specificity, and the rate, timing and number of applications of insecticide made. For instance, insect pests that survive in large populations and breed quickly are at greater advantage of evolving insecticide, especially when insecticides are misused or over-used.

General strategy for insecticide resistance management: The best strategy to avoid insecticide resistance is prevention and including insecticide resistance management tactics as part of a larger integrated pest management (IPM) approach.

1) Monitor pests: Monitor insect population development in fields to determine if and when control measures are warranted. Monitor and consider natural enemies when making control decisions. After treatment, continue monitoring to assess pest populations and their control.

2) Focus on AESA. Insecticides should be used only as a last resort when all other non-chemical management options are exhausted and P: D ratio is above 2: 1.

3) Take an integrated approach to managing pests. Use as many different control measures as possible. Select insecticides with care and consider the impact on future pest populations and the environment. Avoid broad-spectrum insecticides when a narrow-spectrum or more specific insecticide will work.

4) Time applications correctly. Apply insecticides when the pests are most vulnerable. Use application rates and intervals recommended by the manufacturer, university insect management specialist, county Extension agent, or crop consultant.

5) Mix and apply carefully. While applying insecticides care should be taken for proper application of insecticides in terms of dose, volume, timing, coverage, using techniques recommended by the manufacturer etc.

6) Alternate different insecticide classes. Avoid the repeated use of the same insecticide, insecticides in the same chemical class, or insecticides in different classes with same mode of action and rotate/alternate insecticide classes and modes of action.

7) Preserve susceptible genes. Preserve susceptible individuals within the target population by providing unsprayed areas within treated fields, adjacent "refuge" fields, or habitat attractions within a treated field that facilitate immigration. These susceptible individuals may outcompete and interbreed with resistant individuals, diluting the resistant genes and therefore the impact of resistance.

VI. COMMON WEEDS



1. Pigweed: *Amaranthus viridis* Hook. F.
(Amaranthaceae)



2. Swine cress: *Coronopus didymus* (L.) Sm.
(Brassicaceae)



3. Black nightshade: *Solanum nigrum* L.
(Solanaceae)



4. Common purselane: *Portulaca oleracea* L. (Portulacaceae)



5. False amaranth: *Digera arvensis* Forssk. (Amaranthaceae)



6. Rabbit/crow foot grass: *Dactyloctenium aegyptium* (L.) Willd (Poaceae)



7. Crabgrass: *Digitaria sanguinalis* (L.) Scop. (Poaceae)



8. Barnyard grass: *Echinochloa crusgalli* (L.) Beauv. (Poaceae)



9. Purple nutsedge: *Cyperus rotundus* L. (Cyperaceae)



10. Flat sedge: *Cyperus iria* L. (Cyperaceae)



11. Lambs quarter: *Chenopodium album* L. (Chenopodiaceae)



12. Scarlet pimpernel: *Anagallis arvensis* L. (Primulaceae)



13. Sweet clover: *Melilotus*



14. Fine leaf fumitory:



15. Corn spurry: *Spergula*

indica (L.) All. (Fabaceae)

Fumaria parviflora Lam.
(Fumariaceae)

arvensis L.
(Caryophyllaceae)



16. Bluegrass: *Poa annua*
L. (Poaceae)



17. Canary grass: *Phalaris minor* Retz. (Poaceae)

VII. DESCRIPTION OF INSECT PESTS

1) Moringa hairy caterpillar:

It is a specific and common pest on drumstick.

Biology:

Adult: It is a large moth with light yellowish brown wings, having faint lines. Female moth lays eggs in clusters on tender parts.

Egg: Egg period is 5-7 days.

Larva: Caterpillar is dirty brown in colour with whitish hairs arising in tufts on small warts. Hairs are poisonous and irritating. Head capsule and thoracic legs are coral red in colour. Larval period is 20-25 days.

Pupa: Larva pupates in soil in an earthen cocoon. Pupal period is 8-10 days.



Larva

Damage symptoms:

- Caterpillars gather in a cluster on the stem of the plants during hot hours of day
- They are active at night, defoliate the tree quickly and collect on the trunk
- The larva feeds on leaves causing defoliation



Hairy caterpillars Source: SK Sain

*For management refer to page number-----

2) Moringa budworm:

Biology:

Egg: Creamy oval eggs laid singly or in groups on flower buds.

Larva: Dirty brown with a prominent mid dorsal stripe, black head and prothoracic shield

Pupa: Pupation takes place in an earthen cocoon in the soil

Adult: Fore and hind wings are light yellowish brown

Nature and symptoms of damage:

- In early stages, the caterpillars are gregarious and scrape the chlorophyll content of leaf lamina giving it a papery white appearance.
- Later they become voracious feeders making irregular holes on the leaves.
- Irregular holes on leaves initially and later skeletonization leaving only veins and petioles
- Heavy defoliation.
- Bored fruits with irregular holes
- Larvae bore into flower buds feeding on the inner contents and cause shedding up to 78 per cent during summer.
- Only one caterpillar is seen in a bud.

*For management refer to page number-----

3) Pod fly:

Biology:

Egg: Cigar shaped, laid in groups on the grooves of tender pods.

Maggot: Cream coloured maggot

Adult: Yellowish fly with red eyes



Adult:

*For management refer to page number-----

4) Leaf caterpillar:

Biology:

Egg: Creamy white oval eggs are laid in clusters on leaves.

Larva: Devoid of prothoracic shield.

Adult: Similar to budworm but bigger in size



Larva



Adult

Damage symptoms:

- Caterpillars feed on the leaves and cause defoliation.

*For management refer to page number-----

5) Bark caterpillar:

Adult: Adult is pale brown moth. Fore and hindwings are cream-white in color. Forewings have brown spots and streaks.



Damage symptoms:

- Zig-zag galleries and silken webbed masses comprising of chewed material and excreta of larva



*For management refer to page number-----

VIII. DESCRIPTION OF DISEASES

1) Damping off:

Disease symptoms:

Disease of nursery beds and young seedlings resulting in reduced seed germination and poor stand of seedlings.

Very high seedling mortality 25-75%

- **Pre-emergence** damping off: Seedlings disintegrate before they come out of soil surface leading to poor seed germination
- **Post-emergence** damping off is characterised by development of disease after seedlings have emerged out of soil but before the stems are lignified
- **Water soaked lesion** formation at collar region
- Infected areas turn **brown and rot**
- Plants shrivel and **collapse** as a result of softening of tissues
- In *Rhizoctonia solani* attack infected stems become hard, thin (**wire stem** symptoms) and infected seedlings topple Disease appear in patches both in nursery and field beds.



Damping off symptom on stem Source: SK Sain

Survival and spread:

Primary: Oospores in soil in case of *Pythium*

Sclerotia in soil in case of *Rhizoctonia*

Secondary: Zoospores through irrigation water in case of *Pythium*

Mycelial growth in soil and sclerotia through irrigation water in case of *Rhizoctonia*

Favourable conditions:

For *Pythium*

- Heavy rainfall
- Excessive and frequent irrigation
- Poorly drained soil and close spacing
- High soil moisture with temp around 25-30°C

For *Rhizoctonia*

- High soil moisture with temp around 30–35°C

*For management refer to page number-----

2) Twig canker:

Disease symptoms:

- The first symptom of the disease is clearing of the veinlets and chlorosis of the leaves.
- The younger leaves may die in succession and the entire may wilt and die in a course of few days.
- Soon the petiole and the leaves droop and wilt.
- In young plants, symptom consists of clearing of veinlet and dropping of petioles.
- The symptoms continue in subsequent leaves.
- At later stage, browning of vascular system occurs.
- Plants become stunted and die.

Survival and spread:

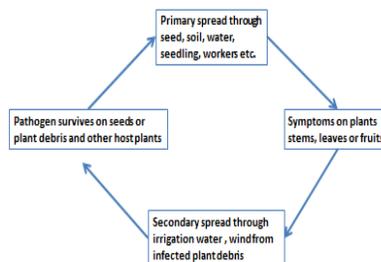
- Soil and implements

Favourable conditions:

- Relatively high soil moisture and soil temperature

Disease cycle

1) Twig canker:



IX. SAFETY PRECAUTIONS

A. During post-harvest storage:

Full leaves (leaflets plus wiry stalks) should be washed carefully with water of drinking quality or clean seawater. If bundle wrapped in moist paper and kept in a cool location they should store for a day. Leaves can last for up to a week, if placed in an airtight container in a cool room or refrigerator. If the leaves dry they will drop their leaflets and lose their value as a food.

X. DO'S AND DON'TS

S. No.	Do's	Don'ts
1.	Deep ploughing is to be done on bright sunny days during the months of May and June. The field should be kept exposed to sun light at least for 2-3 weeks	Do not plant or irrigate the field after ploughing, at least for 2-3 weeks, to allow desiccation of weed's bulbs and/or rhizomes of perennial weeds.
2.	Adopt crop rotation.	Avoid monocropping.
3.	Grow only recommended varieties.	Do not grow varieties not suitable for the season or the region.
4	Sow early in the season	Avoid late sowing as this may lead to reduced yields and incidence of white grubs and diseases.
5	Always treat the seeds with approved chemicals/bioproducts for the control of seed borne diseases/pests.	Do not use seeds without seed treatment with biopesticides/chemicals.
6.	Sow in rows at optimum depths under proper moisture conditions for better establishment.	Do not sow seeds beyond 5-7 cm depth.
7.	Apply only recommended herbicides at recommended dose, proper time, as appropriate spray solution with standard equipment along with flat fan or flat jet nozzles.	Pre-emergent as well as soil incorporated herbicides should not be applied in dry soils. Do not apply herbicides along with irrigation water or by mixing with soil, sand or urea.
8.	Maintain optimum and healthy crop stand which would be capable of competing with weeds at a critical stage of crop weed competition	Crops should not be exposed to moisture deficit stress at their critical growth stages.
9	Use NPK fertilizers as per the soil test recommendation.	Avoid imbalanced use of fertilizers.
10	Use micronutrient mixture after sowing based test recommendations.	Do not apply any micronutrient mixture after sowing without test recommendations.
11	Conduct AESA weekly in the morning preferably before 9 a.m. Take decision on management practice based on AESA and P: D ratio only.	Do not take any management decision without considering AESA and P: D ratio
12	Install pheromone traps at appropriate period.	Do not store the pheromone lures at normal room temperature (keep them in refrigerator).
13	Release parasitoids only after noticing adult moth catches in the pheromone trap or as pheromone trap or as per field observation	Do not apply chemical pesticides within seven days of release of parasitoids.
14	Apply NPV, if available, at recommended dose when a large number of egg masses and early instar larvae are noticed. Apply NPV only in the evening hours after 5 pm.	Do not apply NPV on late instar larva and during day time.

15	In case of pests which are active during night spray recommended biopesticides/chemicals at the time of their appearance in the evening.	Do not spray pesticides at midday since, most of the insects are not active during this period.
16	Spray pesticides thoroughly to treat the undersurface of the leaves, particularly for sucking pests	Do not spray pesticides only on the upper surface of leaves.
17	Apply short persistent pesticides to avoid pesticide residue in the soil and produce.	Do not apply pesticides during preceding 7 days before harvest.
18	Follow the recommended procedure of trap crop technology.	Do not apply long persistent pesticides on trap crop, otherwise it may not attract the pests and natural enemies.

XI. BASIC PRECAUTIONS IN PESTICIDES USAGE

A. Purchase

1. Purchase only just required quantity e.g. 100, 250, 500, 1000 g/ml for single application in specified area.
2. **Do not** purchase leaking containers, loose, unsealed or torn bags; **Do not** purchase pesticides without proper/approved labels.
3. While purchasing insist for invoice/bill/cash memo

B. Storage

1. Avoid storage of pesticides in house premises.
2. Keep only in original container with intact seal.
3. **Do not** transfer pesticides to other containers; **Do not** expose to sunlight or rain water; **Do not** store weedicides along with other pesticides.
4. Never keep them together with food or feed/fodder.
5. Keep away from reach of children and livestock.

C. Handling

1. Never carry/ transport pesticides along with food materials.
2. Avoid carrying bulk pesticides (dust/granules) on head shoulders or on the back.

D. Precautions for preparing spray solution

1. Use clean water.
2. Always protect your nose, eyes, mouth, ears and hands.
3. Use hand gloves, face mask and cover your head with cap.
4. Use polythene bags as hand gloves, handkerchiefs or piece of clean cloth as mask and a cap or towel to cover the head (Do not use polythene bag contaminated with pesticides).
5. Read the label on the container before preparing spray solution.
6. Prepare the spray solution as per requirement
7. **Do not** mix granules with water; **Do not** eat, drink, smoke or chew while preparing solution
8. Concentrated pesticides must not fall on hands etc. while opening sealed container. Do not smell pesticides.

9. Avoid spilling of pesticides while filling the sprayer tank.
10. The operator should protect his bare feet and hands with polythene bags

E. Equipment

1. Select right kind of equipment.
2. **Do not** use leaky and defective equipment
3. Select right kind of nozzles
4. **Do not** blow/clean clogged nozzle with mouth. Use old tooth brush tied with the sprayer and clean with water.
5. **Do not** use same sprayer for weedicide and insecticide.

F. Precautions for applying pesticides

1. Apply only at recommended dose and dilution
2. **Do not** apply on hot sunny day or strong windy condition; **Do not** apply just before the rains and after the rains; **Do not** apply against the windy direction
3. Emulsifiable concentrate formulations should not be used for spraying with battery operated ULV sprayer
4. Wash the sprayer and buckets etc. with soap water after spraying
5. Containers buckets etc. used for mixing pesticides should not be used for domestic purpose
6. Avoid entry of animals and workers in the field immediately after spraying
7. Avoid tank mixing of different pesticides

G. Disposal

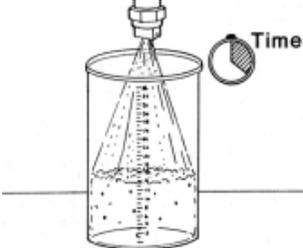
1. Left over spray solution should not be drained in ponds or water lines etc. throw it in barren isolated area if possible
2. The used/empty containers should be crushed with a stone/stick and buried deep into soil away from water source.
3. Never reuse empty pesticides container for any other purpose.

XII. PESTICIDE APPLICATION TECHNIQUES

Equipment			
Category A: Stationary, crawling pest/disease			
Vegetative stage i) For crawling and soil borne pests	Insecticides and fungicides	<ul style="list-style-type: none"> • Lever operated knapsack sprayer (droplets of big size) • Hollow cone nozzle @ 35 to 40 psi • Lever operating speed = 15 to 20 strokes/min 	
ii) For small sucking leaf borne pests		<p>or</p> <ul style="list-style-type: none"> • Motorized knapsack sprayer or mist blower (droplets of small size) • Airblast nozzle • Operating speed: 2/3rd throttle 	

Reproductive stage	Insecticides and fungicides	<ul style="list-style-type: none"> Lever operated knapsack sprayer (droplets of big size) Hollow cone nozzle @ 35 to 40 psi Lever operating speed = 15 to 20 strokes/min 	
Category B: Field flying pest/airborne pest			
Vegetative stage	Insecticides and fungicides	<ul style="list-style-type: none"> Motorized knapsack sprayer or mist blower (droplets of small size) Airblast nozzle Operating speed: 2/3rd throttle Or <ul style="list-style-type: none"> Battery operated low volume sprayer (droplets of small size) Spinning disc nozzle 	
Reproductive stage (Field Pests)			
Mosquito/ locust and spatial application (migratory Pests)	Insecticides and fungicides	<ul style="list-style-type: none"> Fogging machine and ENV (exhaust nozzle vehicle) (droplets of very small size) Hot tube nozzle 	
Category C: Weeds			
Post-emergence application	Weedicide	<ul style="list-style-type: none"> Lever operated knapsack sprayer (droplets of big size) Flat fan or floodjet nozzle @ 15 to 20 psi Lever operating speed = 7 to 10 strokes/min 	 
Pre-emergence application	Weedicide	<ul style="list-style-type: none"> Trolley mounted low volume sprayer (droplets of small size) Battery operated low volume sprayer (droplets of small size) 	

XIII. OPERATIONAL, CALIBRATION AND MAINTENANCE GUIDELINES IN BRIEF

1.	For application rate and dosage see the label and leaflet of the particular pesticide.	 
2.	It is advisable to check the output of the sprayer (calibration) before commencement of spraying under guidance of trained person.	 
3.	Clean and wash the machines and nozzles and store in dry place after use.	  
4.	It is advisable to use protective clothing, face mask and gloves while preparing and applying pesticides. Do not apply pesticides without protective clothing and wash clothes immediately after spray application.	 
5.	Do not apply in hot or windy conditions.	 

6.	Operator should maintain normal walking speed while undertaking application.	
7.	Do not smoke, chew or eat while undertaking the spraying operation	
8.	Operator should take proper bath with soap after completing spraying	
9.	Do not blow the nozzle with mouth for any blockages. Clean with water and a soft brush.	

XIV. REFERENCES

- <http://tnau.ac.in/eagri/eagri50/ENTO331/lecture26/lec026.pdf>
- <http://www.cabi.org>
- <http://agritech.tnau.ac.in/>
- Satti, A. A., Nasr, O. El-Hag, Fadelmula, A., Eshag Ali, F. 2013. New record and preliminary bioecological studies of the leaf caterpillar, *Noorda blitealis* Walker (Lepidoptera: Pyralidae) in Sudan. International Journal of Science and Nature 4: 57-62.
- <http://zsi.gov.in/checklist/Gaurav-Checklist.pdf>